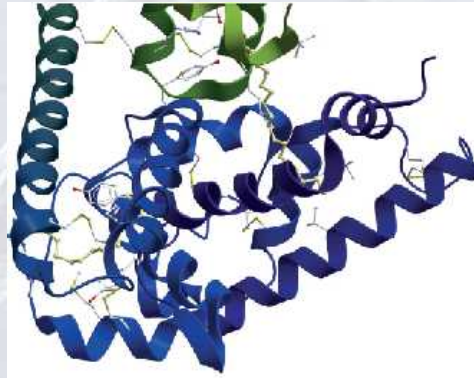




# FORMAMP

INNOVATIVE NANOFORMULATION OF ANTIMICROBIAL PEPTIDES



Helena Bysell (Ph D, Project Manager)  
SP Chemistry, Materials and Surfaces- Life Science

*Early discovery of new antibiotics workshop  
12-13 January 2017*

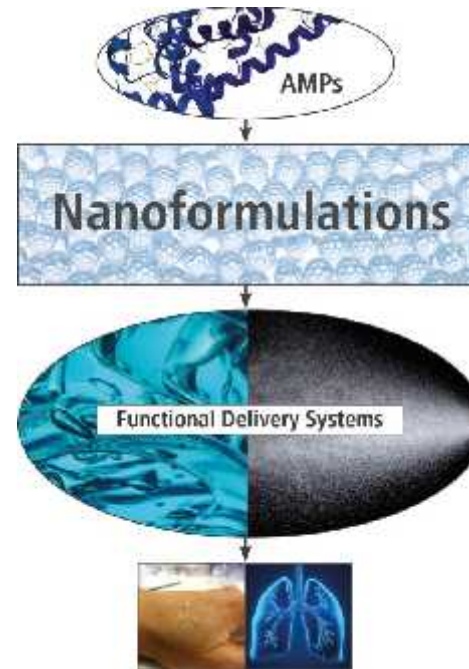
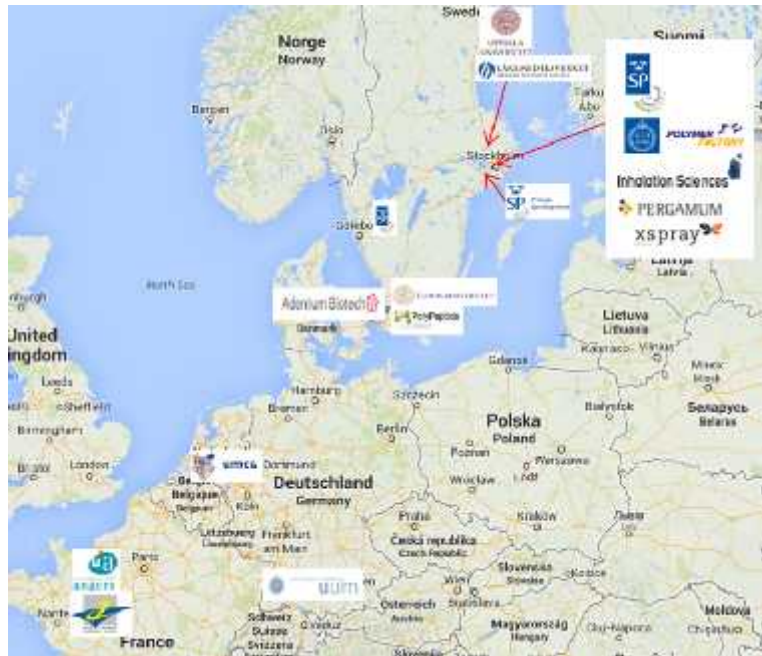




# FORMAMP Innovative Nanoformulation of Antimicrobial peptides

**Vision:** To reduce the alarming progress of multidrug-resistant bacteria

**Mission:** To develop new sustainable strategies for treatment of infectious diseases



## Facts:

Project duration: 2013-2017

Budget: 10.5 MEuro,

EU contribution 8 MEuro

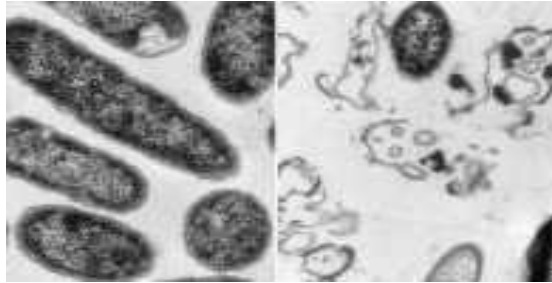
16 partners from 5 countries

Coordinator:

[helena.bysell@sp.se](mailto:helena.bysell@sp.se)

[www.formampproject.com](http://www.formampproject.com)

# Antimicrobial peptides

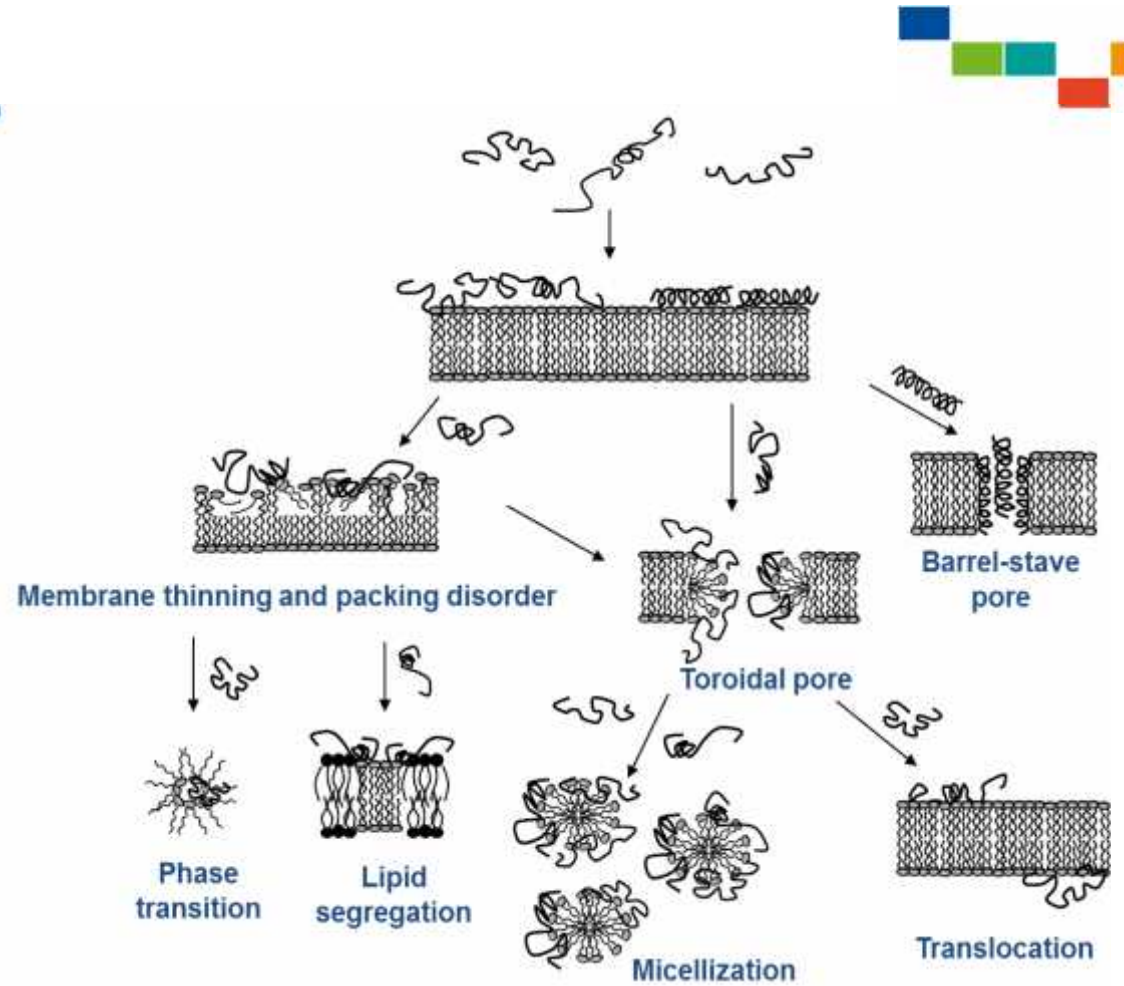


*J Biol Chem*, 2005, 280, 34832

- Fast and non-specific mechanism of action
- Bacteria not as prone to develop high level resistance

## Efficiency and MoA influenced by

- Size
- Conformation
- Net charge
- Charge distribution
- Hydrophobicity



*Ringstad, Uppsala University thesis, 2009*

# AMPs in research and development

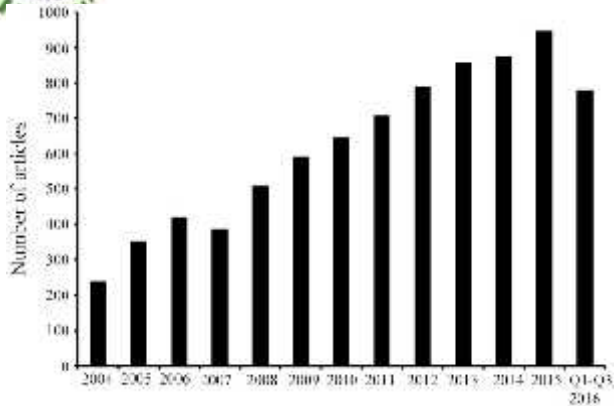


TABLE 1 | Selected AMPs in clinical phase of development.

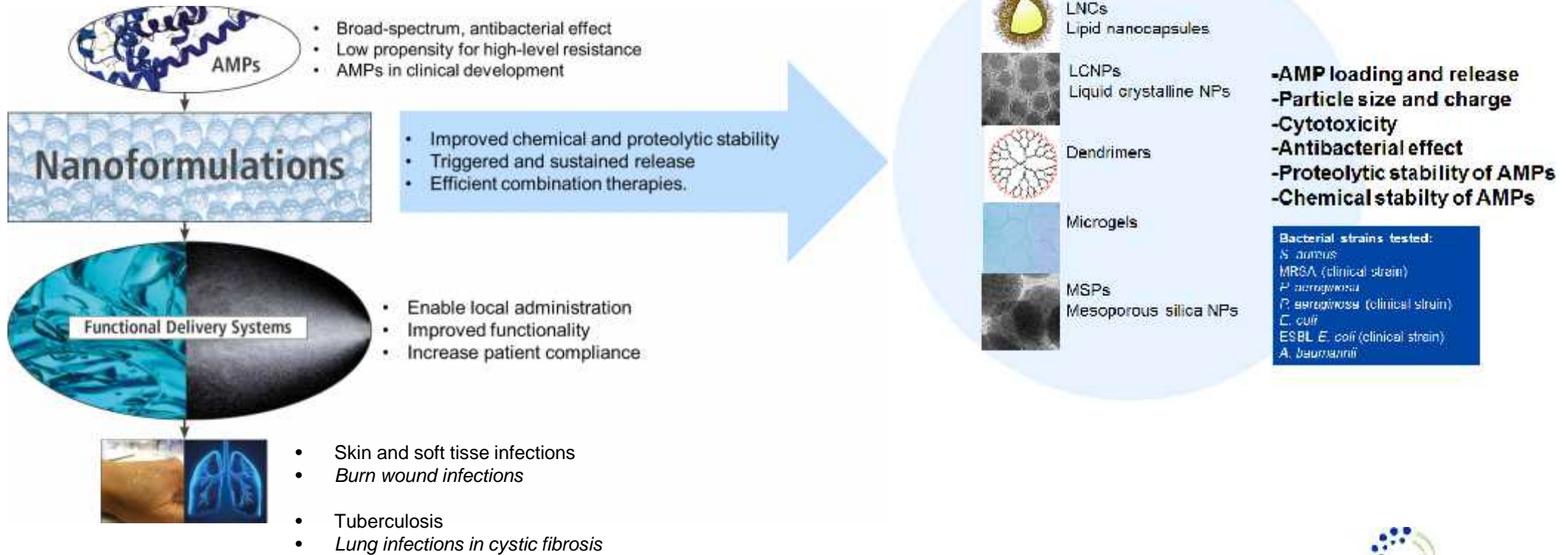
| AMP                | Description                                       | Phase     | Indication  | Administration   | Clinical trial identifier if available |
|--------------------|---|-----------|---|--|--|
| Resiganan (MSH 74) | Analog of mappinir (skin of African clawed frog)  | Phase II  | Infected diabetic foot ulcers   | Topical cream  | NCT00568304, NCT01668313               |
| Omigorel           | Derived from indolicidin (bacteria)               | Phase III | Cellulite infections and abscesses  | Topical gel  | NCT01231159, NCT01784139               |
| Lytlear (LTX 100)  | Synthetic antimicrobial lipopeptides              | Phase II  | Uncomplicated Gram-positive skin infections, impetigo, and nasal colonization with <i>S. aureus</i>   | Topical hydrogel   | NCT01223222, NCT01803125, NCT01158235  |
| hLF1-11            | Derived from lactoferrin (human)                  | Phase II  | Bacteraemia and fungal infections in immunocompromised haematopoietic stem cell transplant recipients | Intravenous treatment (in saline)  | NCT00609938                            |
| Novozimin (NF-213) | Derived from nisin (bacteria)                     | Phase I   | Osteomyelitis (fungal nail infection)   | Topical (medicated) treatment  |  |
| UZEN-002           | Limonc octamer derived from $\alpha$ -MSH (human) | Phase II  | Vaginal candidiasis   | Vaginal gel  |  |
| LL-37              | LL-37 (human)                                     | Phase II  | Hard-to-heal venous leg ulcers  | Polyvinyl alcohol-based solution for administration in the wound bed                   |  |
| PXL01              | Derived from lactoferrin (human)                  | Phase I   | Prevention of post-surgical adhesion formation in the surgical cavity                                 | Hyaluronic acid-based hydrogel for peritoneal adhesion prevention at the surgical site | NCT01022242                            |
| Resiganan (R3-367) | Derived from protegrin 1 (porcine leukocytes)     | Phase II  | Oral mucositis in patients receiving radiotherapy for head and neck malignancy                        | Oral solution  | NCT00223772                            |
|                    |   | Phase I   | Oral candidiasis in HIV seropositive patients   | Mouthrinse   | NCT00609971                            |

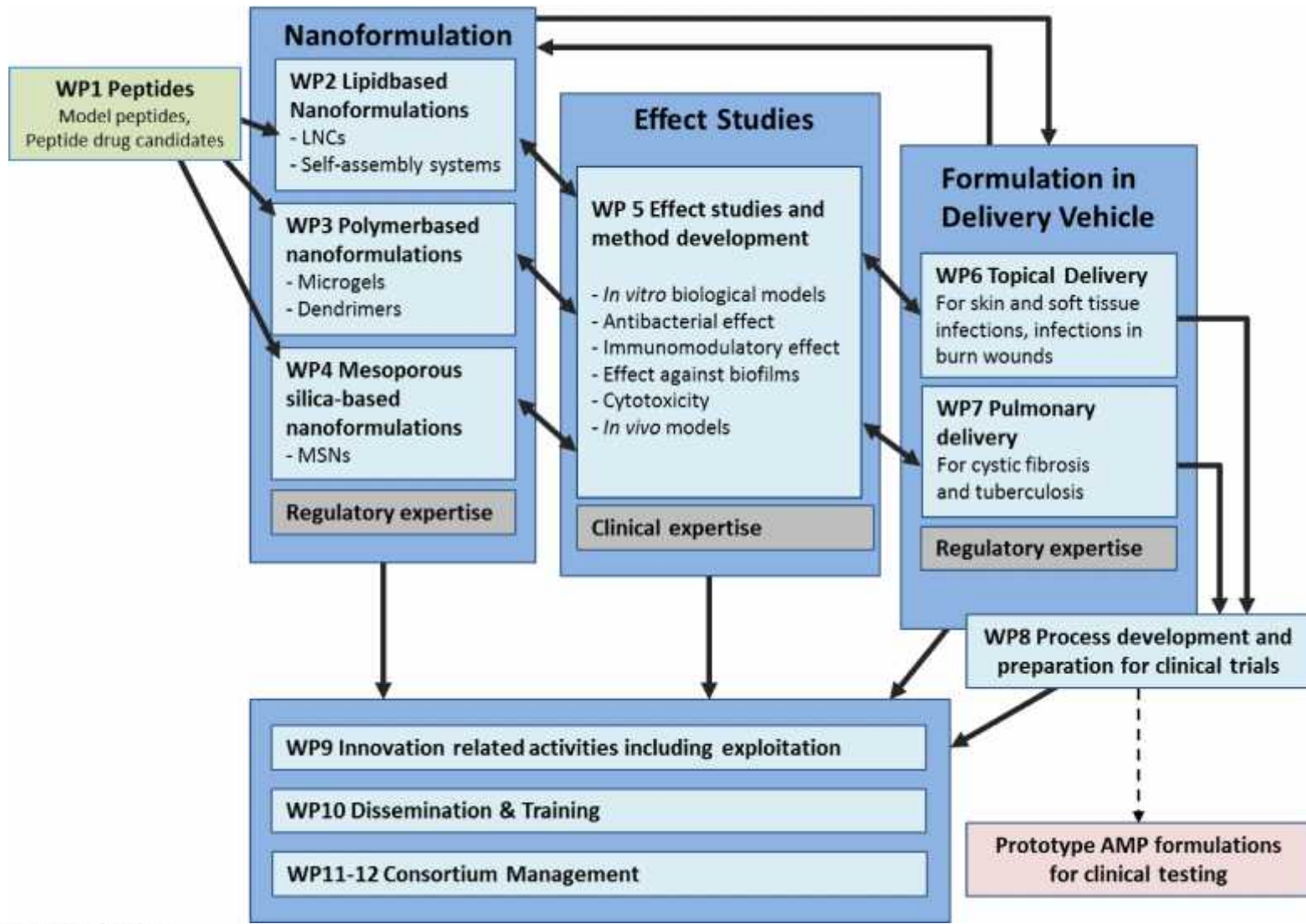
Problems with:  
 -Stability  
 -Toxicity  
 -High cost

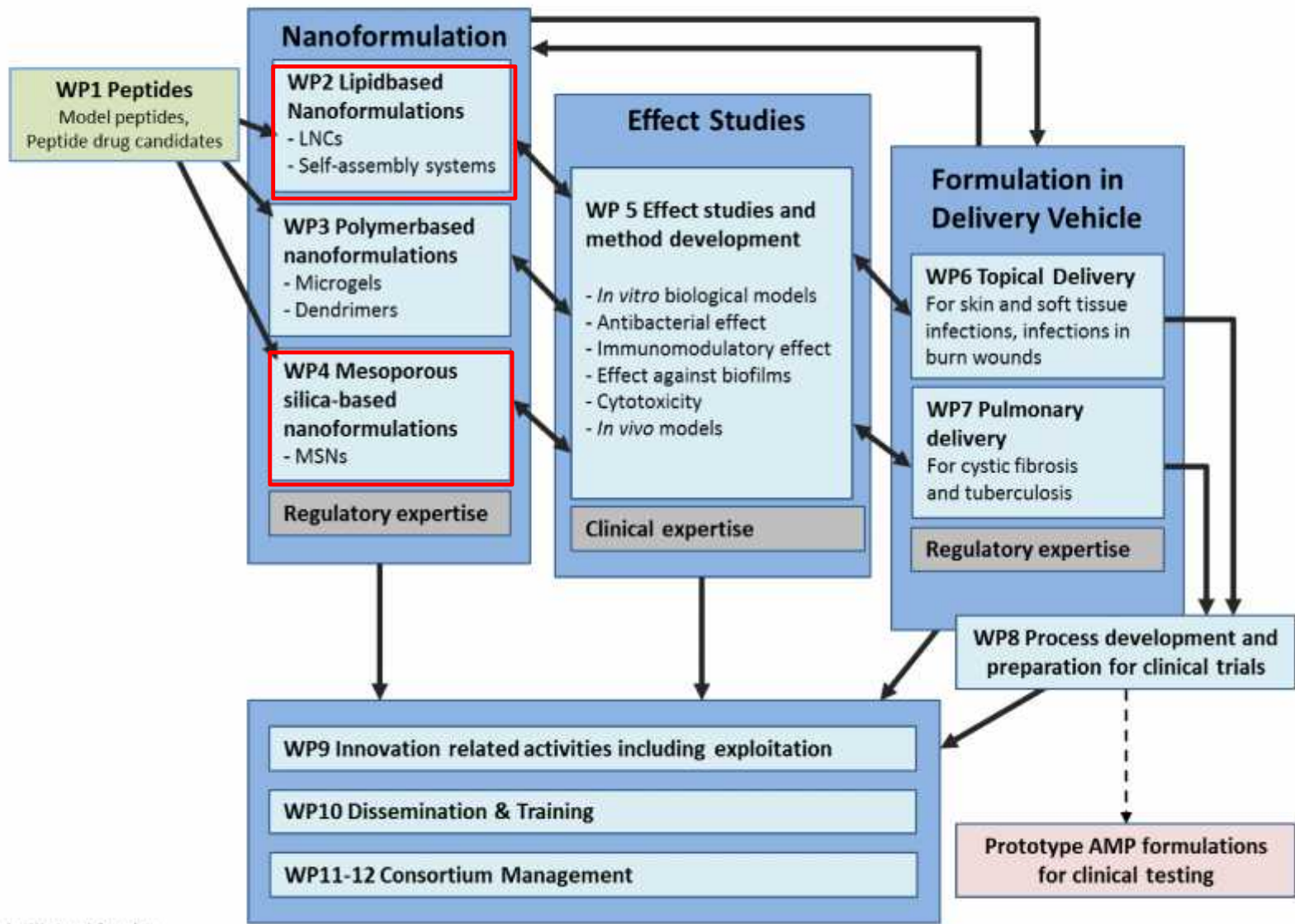
## Formulation



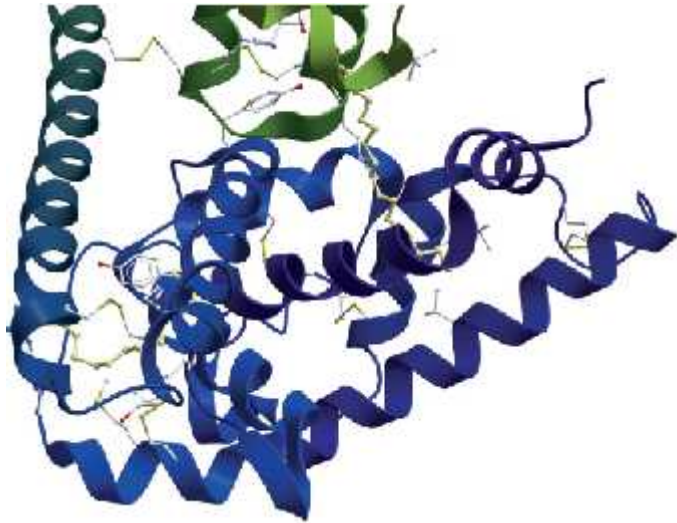
# FORMAMP concept







## Results -Peptides



-5 clinical AMP candidates have been selected and synthesized in mg-g-scale.

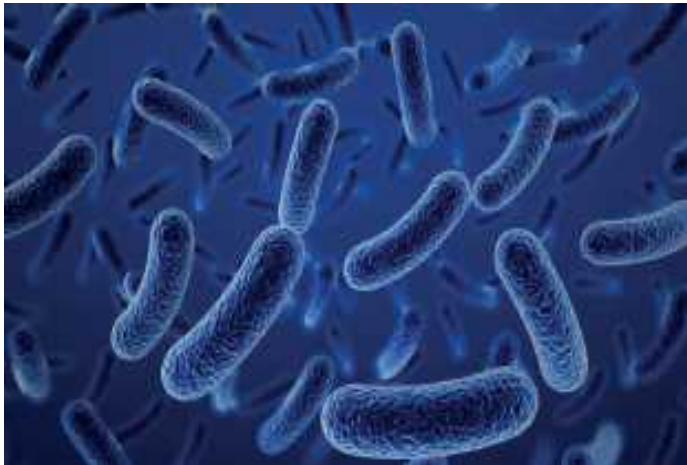
-A peptide effective against *Mycobacterium tuberculosis*, both intracellular and extracellular, has been identified, synthesized and is currently evaluated *in vivo*.

-Target product profiles (TPPs) for the different peptides have been developed.

-Analytical methods have been developed and optimized



## Results –AMP and nanocarriers



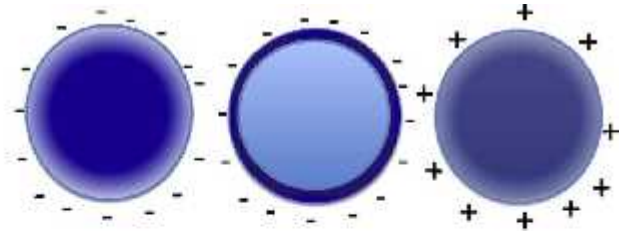
-AMPs can be successfully loaded into the different nanocarrier systems and strategies for optimizing the loading and release are being evaluated.

-More than 400 peptide-carrier combinations have been evaluated *in vitro*. Results show that the antibacterial effect is preserved in 80% of the cases for encapsulated AMPs and also enhanced for 10% of the peptide-carrier combinations.

-AMPs can be protected against proteolytic degradation in selected nanocarriers.

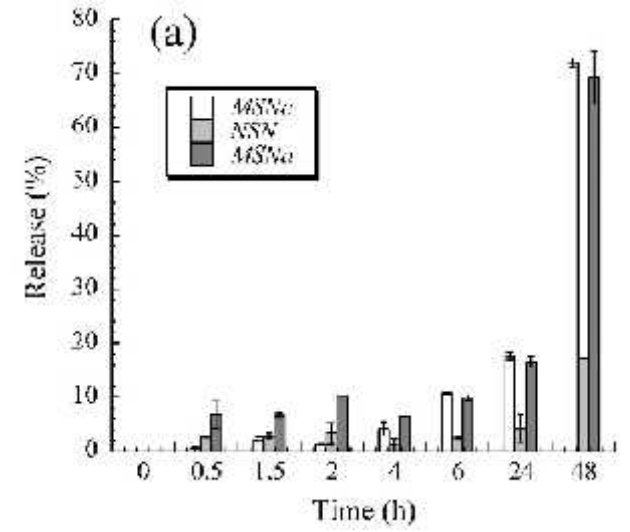
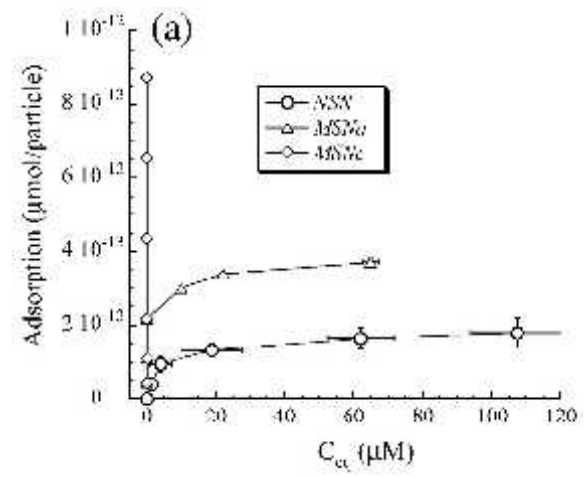
-The majority of AMP-loaded particles display good results with respect to cytotoxicity.

# Formulation of AMPs in mesoporous silica



MSNc (-)      NSN (-)      MSNa (+)

|                       |         |          |          |
|-----------------------|---------|----------|----------|
| Peptide load          | +++++   | +        | +        |
| Location              | Pore    | Surface  | Pore     |
| Antimicrobial entity  | Peptide | Particle | Particle |
| Proteolytic stability | +       | -        | -        |
| Toxicity              | -       | -        | +        |



-Surface charge and surface area strongly influences loading and release of LL-37 and thereby membrane interactions and antimicrobial effect  
 -Peptide localization correlates to proteolytic stability

# Formulation of AMPs in mesoporous silica



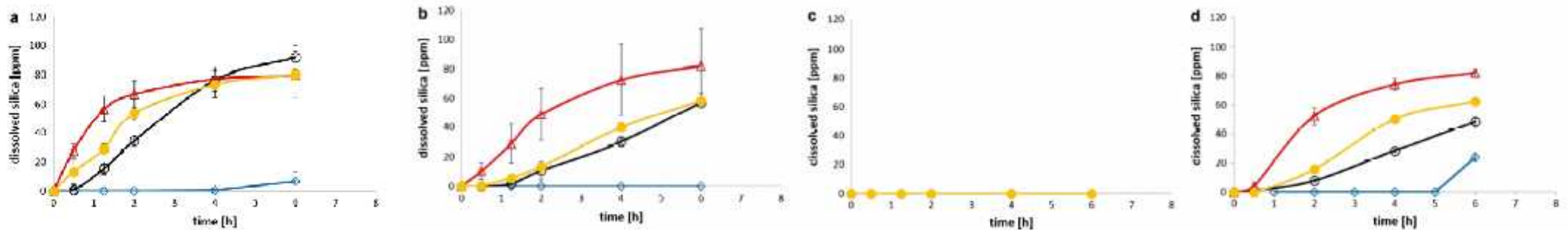
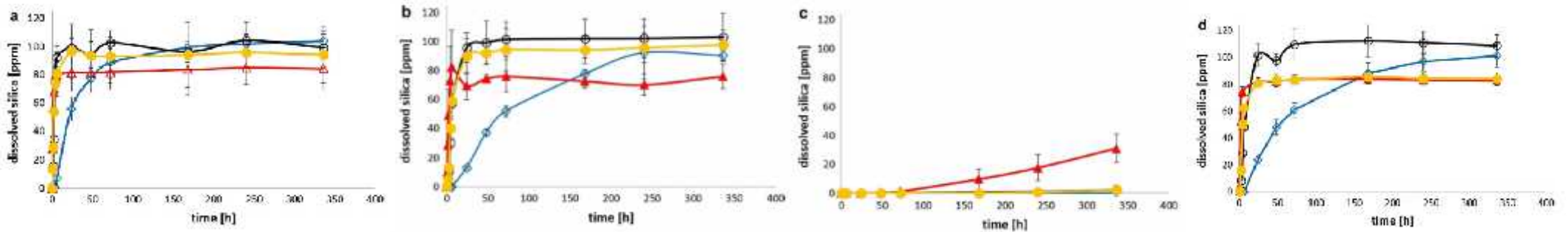
Dissolution kinetics of mesoporous silica nanoparticles in different simulated body fluids

Lung Fluid (SLF)

Body Fluid (SBF)

Gastric Fluid (SGF)

PBS

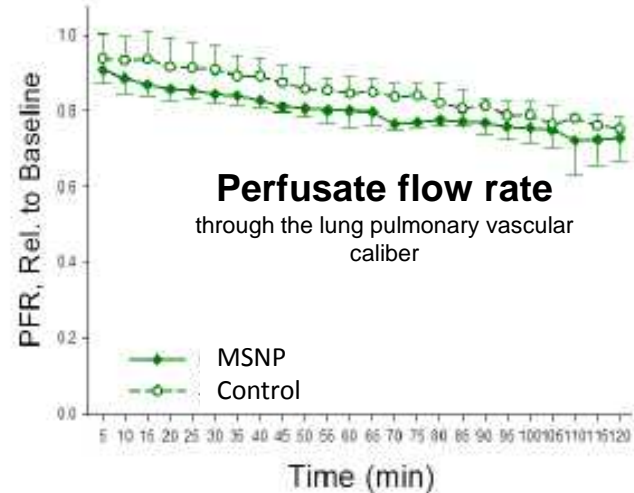
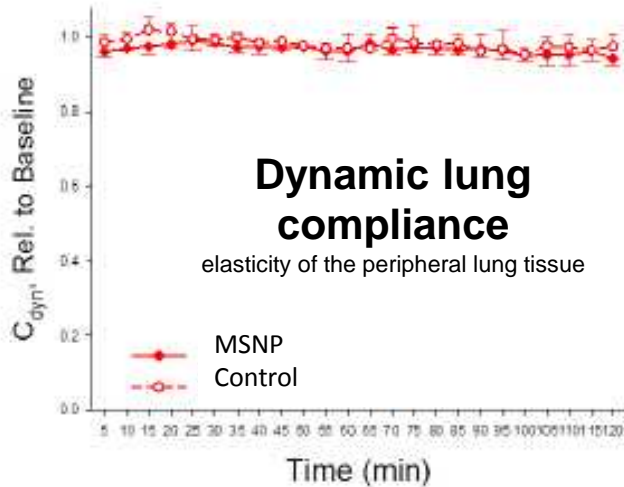
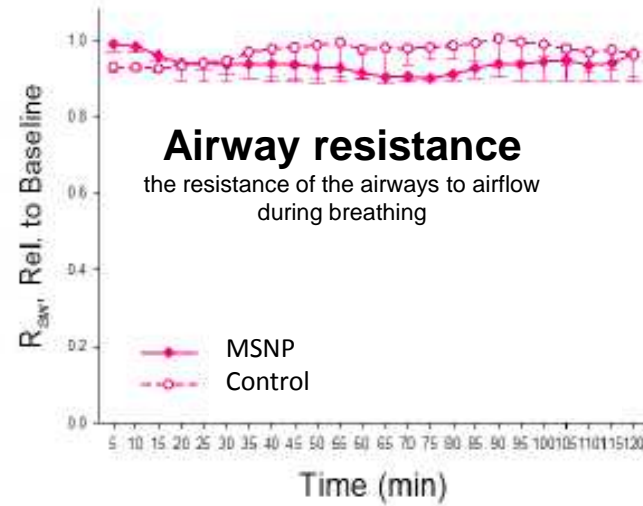
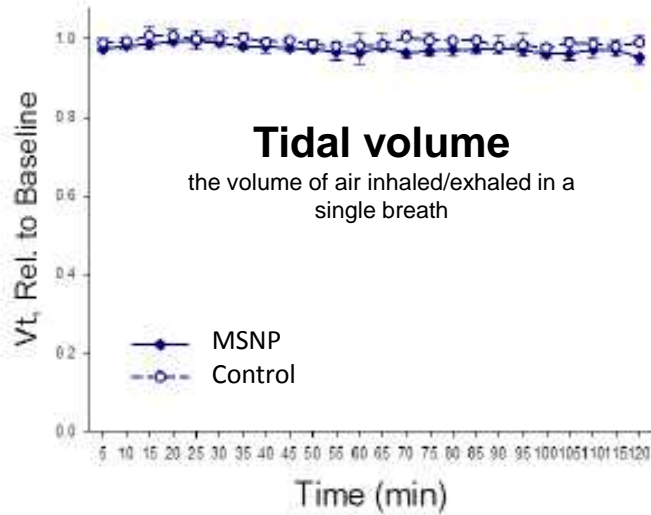


- non-porous 200 nm
- mesoporous 80 nm
- mesoporous 200 nm
- mesoporous 1500 nm

# Formulation of AMPs in mesoporous silica



Ex vivo safety- physiological lung-function variables



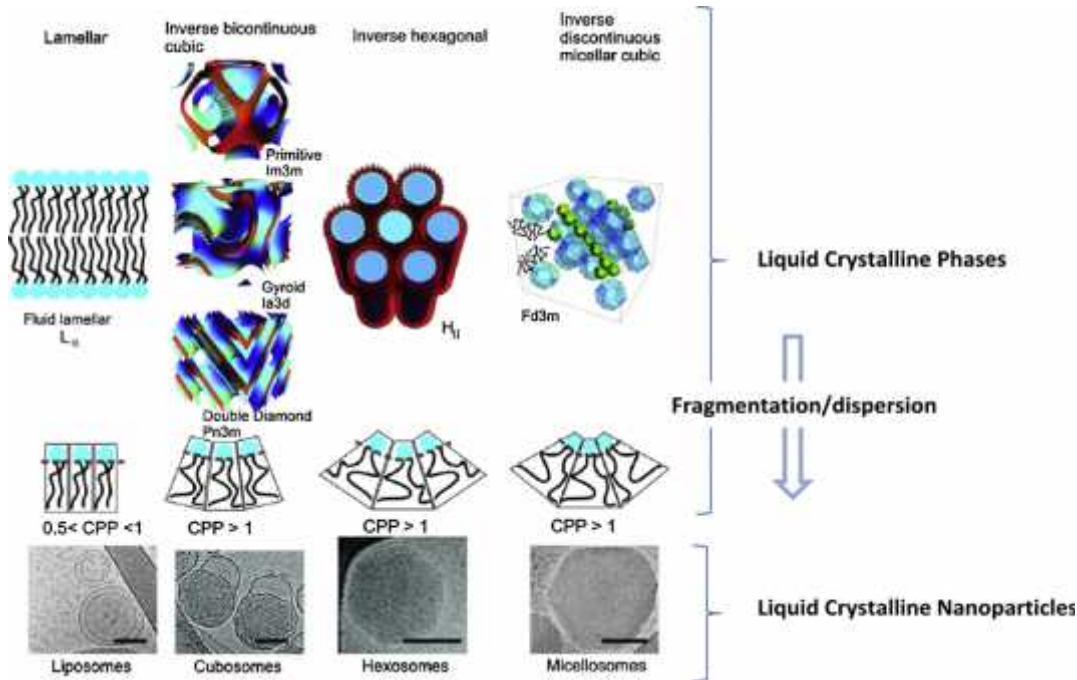
No acute lung toxicity was thus observed in the rat lungs exposed ex vivo to a high dose of particles



# Formulation of AMPs in lipid nanoformulations



## Liquid crystalline nanoparticles (LCNPs)



Mulet et al., 2013

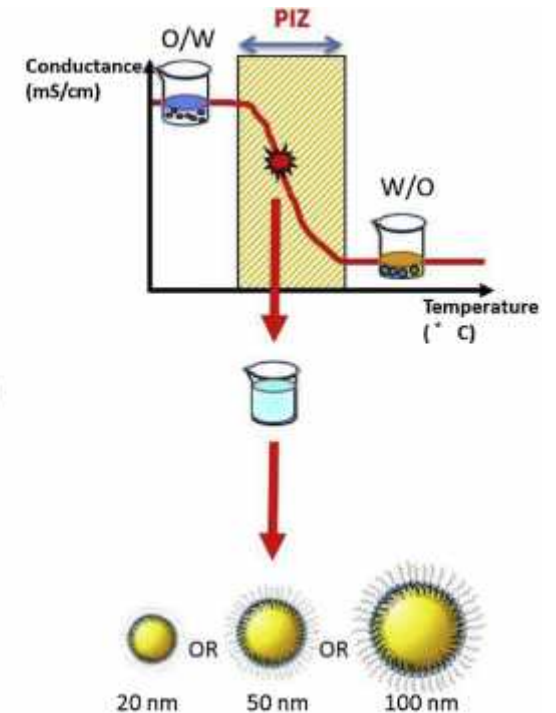
## Lipidic nanocapsules (LNCs)

1- Mix of the components

2- Temperature cycles (60 to 90°C)

3- Rapid cooling in PIZ (water 4°C)

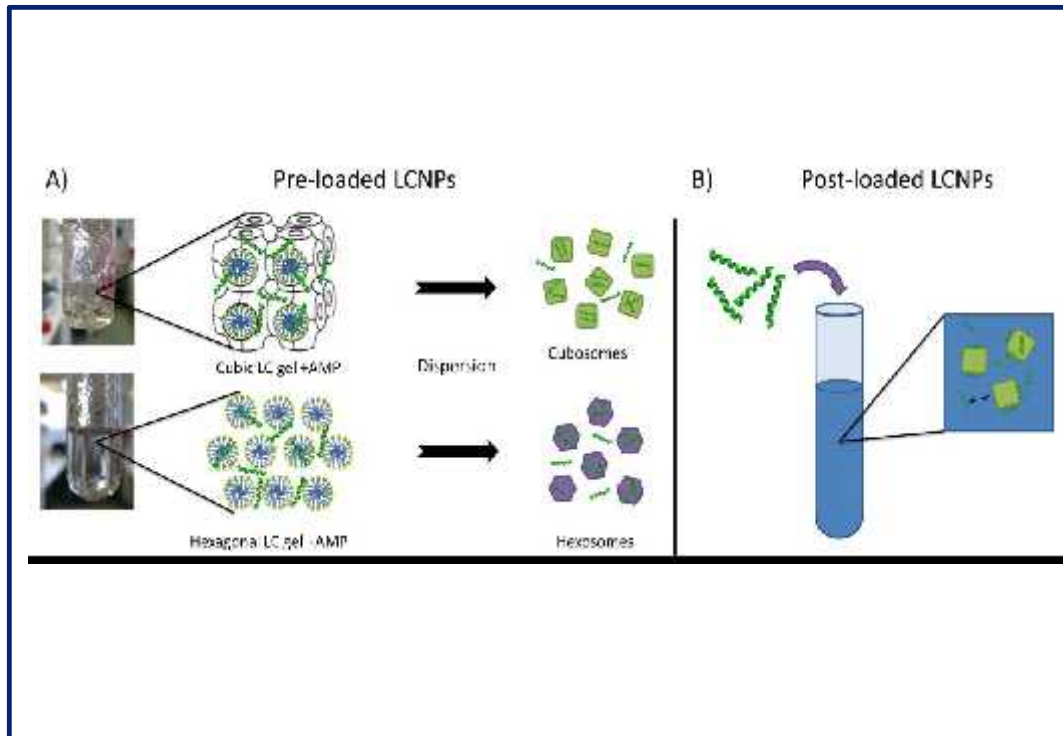
4- Stirring



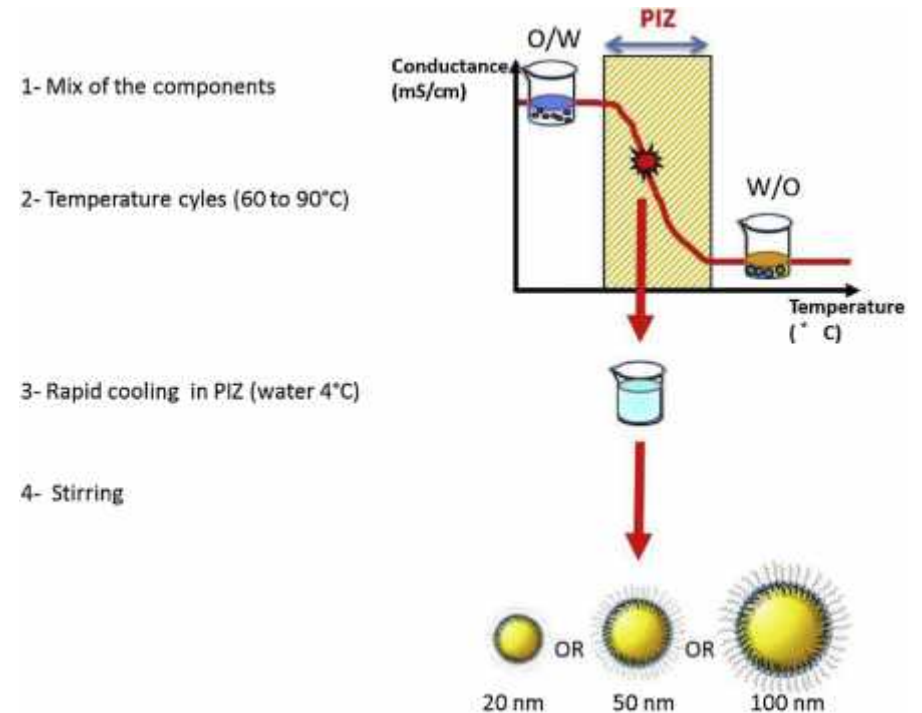
# Formulation of AMPs in lipid nanoformulations



## Liquid crystalline nanoparticles (LCNPs)

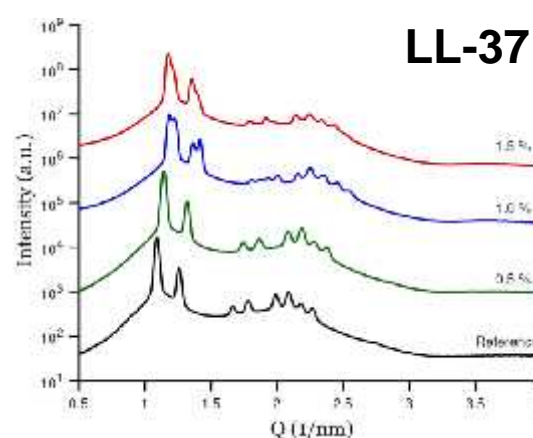
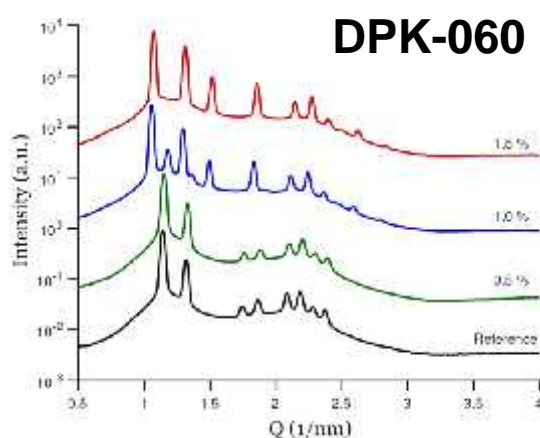
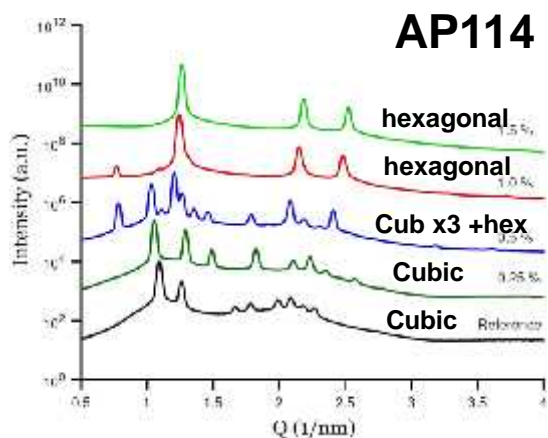


## Lipidic nanocapsules (LNCs)



# Formulation of AMPs in lipid nanoformulations

-Physicochemical characterization- phase behavior



- Cubic → hexagonal
- Increased negative curvature (Ia3d→Hex)

- Cubic → new cubic
- Decreased negative curvature (Ia3d→Pn3m)

- Cubic → same cubic
- No change in curvature (Ia3d)

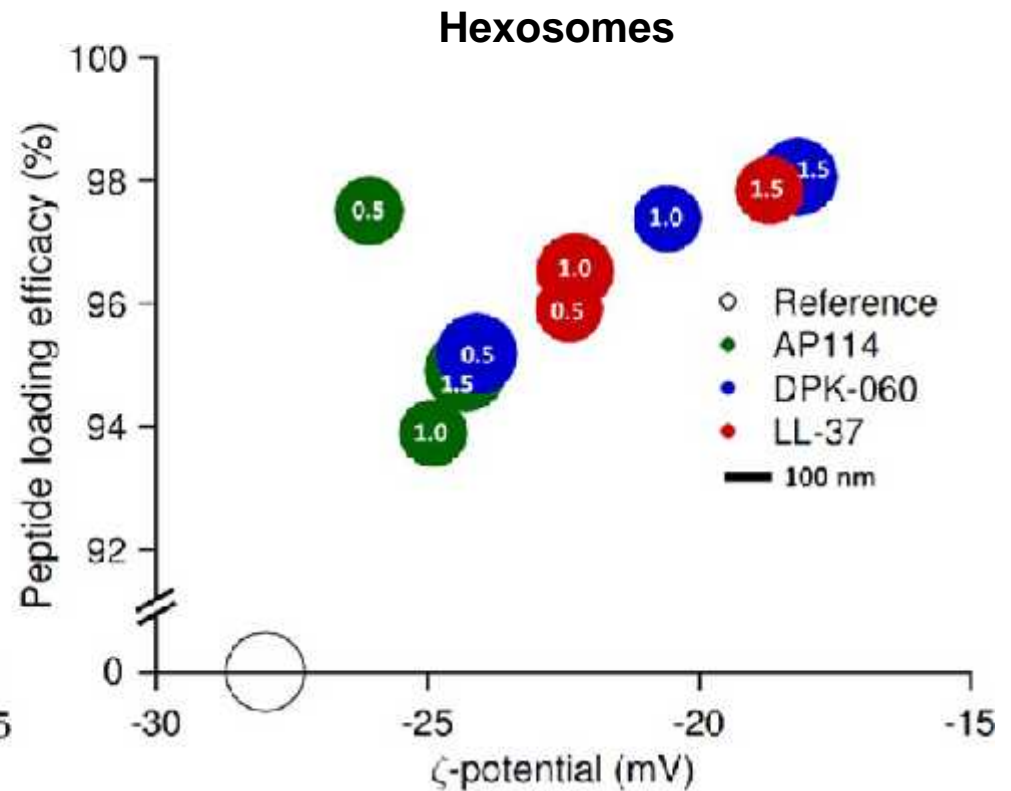
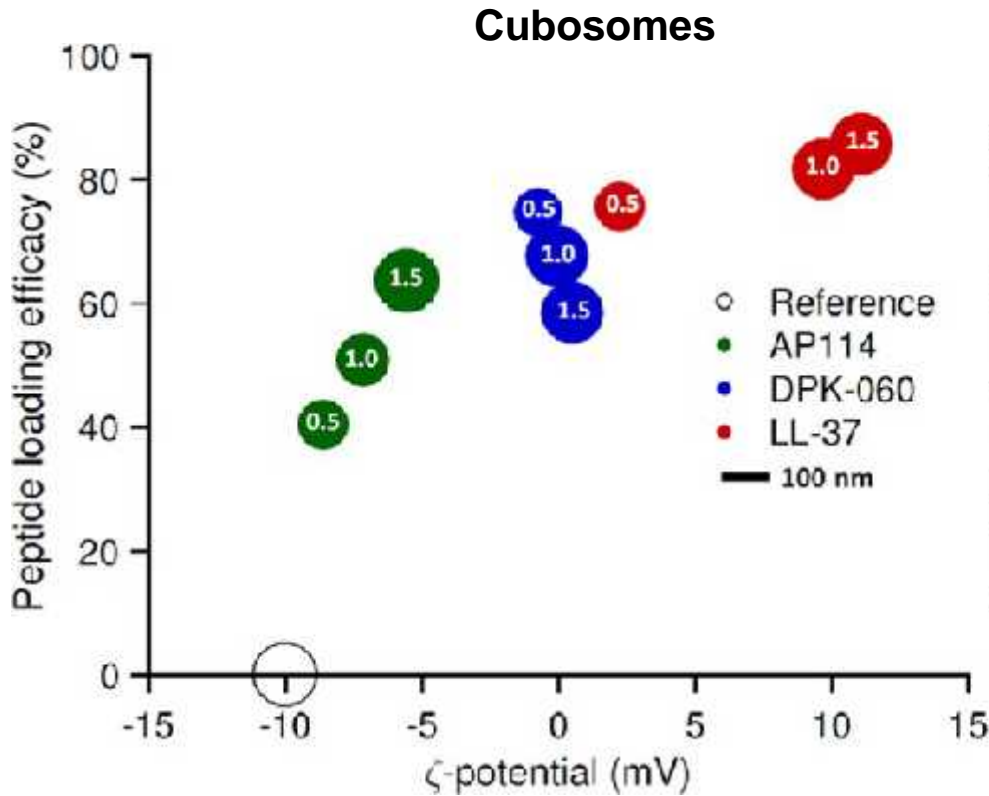
*No phase transitions for hexagonal gels*

|                  |                         |
|------------------|-------------------------|
| Hydrophobicity   | AP114 > LL-37 > DPK-060 |
| Net charge       | DPK-060 > LL-37 > AP114 |
| Molecular weight | LL-37 ~ AP114 > DPK-060 |



# Formulation of AMPs in lipid nanoformulations

-Physicochemical characterization- particle size and charge, peptide loading

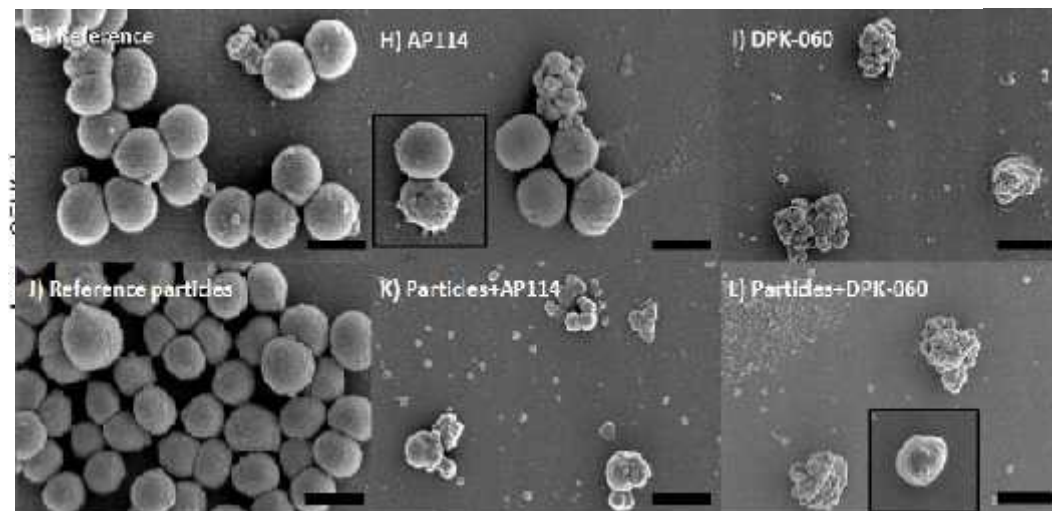
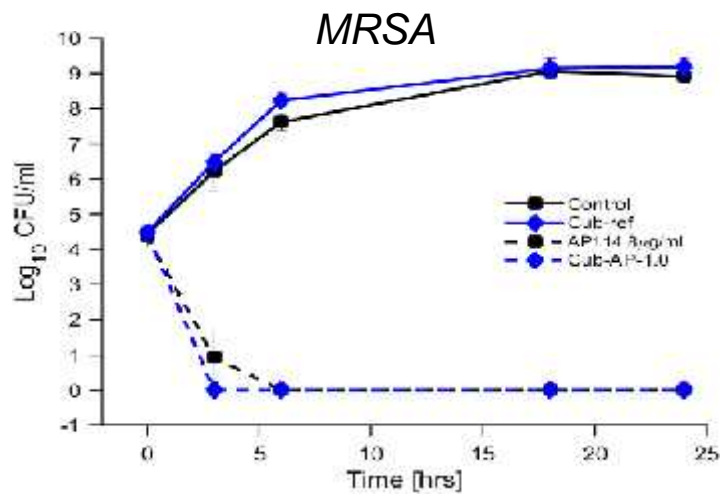




# Formulation of AMPs in lipid nanoformulations

## -Antibacterial effect

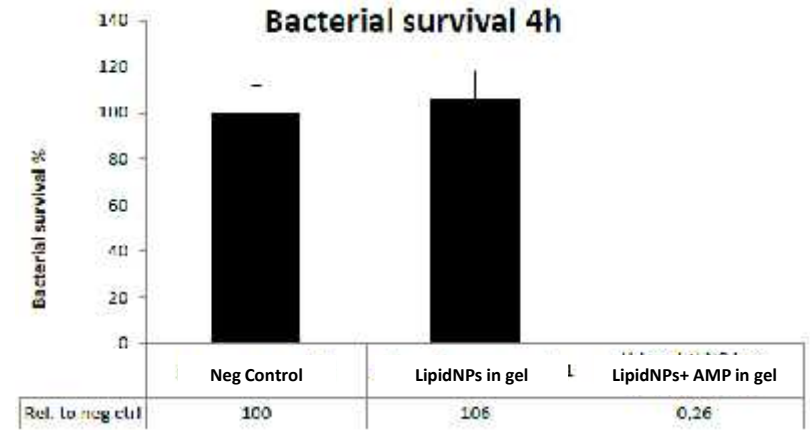
| Bacterial strain                     | MIC values ( $\mu\text{g/mL}$ ) |           |            |         |           |            |       |           |            |
|--------------------------------------|---------------------------------|-----------|------------|---------|-----------|------------|-------|-----------|------------|
|                                      | AP114                           |           |            | DPK-060 |           |            | LL-37 |           |            |
|                                      | UF                              | Pre (GMO) | Post (GMO) | UF      | Pre (GMO) | Post (GMO) | UF    | Pre (GMO) | Post (GMO) |
| <i>S. aureus</i>                     | 8                               | 8         | 8-16       | 4       | 2-4       | 1-2        | 8-16  | >16       | >16        |
| MRSA                                 | 4                               | 4         | 4          | 4       | 2-4       | 2          | 8-16  | >16       | >16        |
| <i>P. aeruginosa</i>                 | -                               | -         | -          | 8       | 16        | 8-16       | 8-16  | >16       | 8-16       |
| <i>P. aeruginosa</i> clinical strain | -                               | -         | -          | 16      | 8-16      | 8          | 8-16  | 8         | 8-16       |
| <i>E. coli</i>                       | -                               | -         | -          | 8       | 2-4       | 4          | 16    | 16        | 8-16       |
| ESBL <i>E. coli</i> clinical strain  | -                               | -         | -          | 4-8     | 2-4       | 2-4        | 16    | >16       | $\geq 16$  |
| <i>A. Baumannii</i>                  | -                               | -         | -          | 4-8     | 16        | 16         | 16    | >16       | $\geq 16$  |



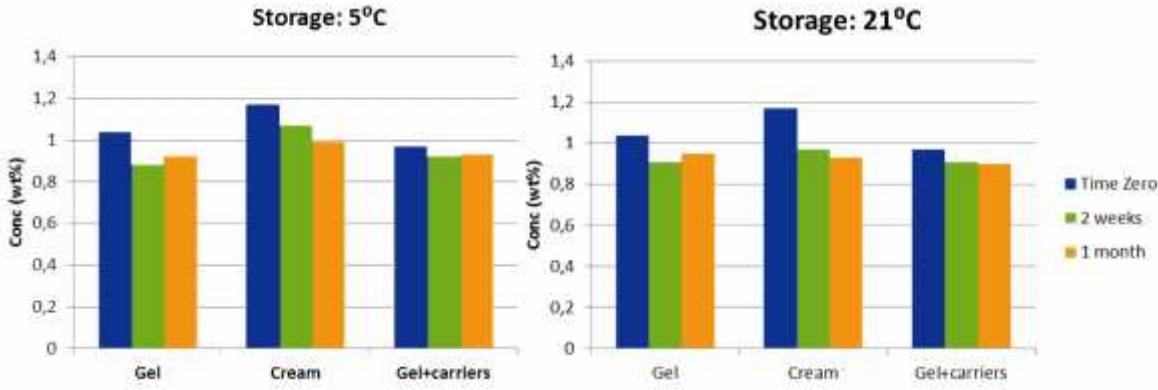


# Formulation of AMPs in lipid nanoformulations

-Ex vivo antibacterial effect and chemical stability of prototype formulations



Stability



## Next steps



- Understand the mechanisms-synergistic effects
- Formulation strategies for biofilms
- Further evaluation of effect and toxicity on refined models (*ex vivo*, *in vivo*).
- Stability studies

**Finding funding for follow up projects!!!**





# Thank you!



[www.formampproject.com](http://www.formampproject.com)

@formamp 



The research in FORMAMP receives funding from the European Union's Seventh Framework Programme (FP7/2007-2013) under grant agreement n° 604182. <http://ec.europa.eu/research>

Martin Andersson  **CHALMERS**  
MAX Lab 

